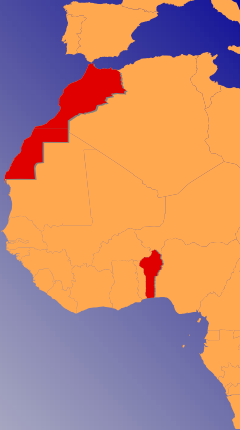




IMPETUS

An integrated approach for efficient and sustainable use of  
fresh water in Western Africa

A5



# *Virological water quality*

Jens Verheyen

Institute of Virology, University of Cologne



University of Cologne

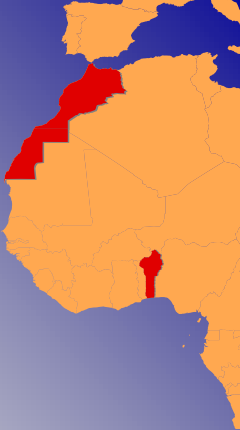


University of Bonn





# *Environmental Virology*



Viruses consist of a genome (RNA or DNA) and several proteins (structure / enzymes) but own no metabolism. For replication viruses use host cells and their organelles.

Viruses are spread to the environment by human fluids (mainly feces, respiratory secrets, vomits, blood).

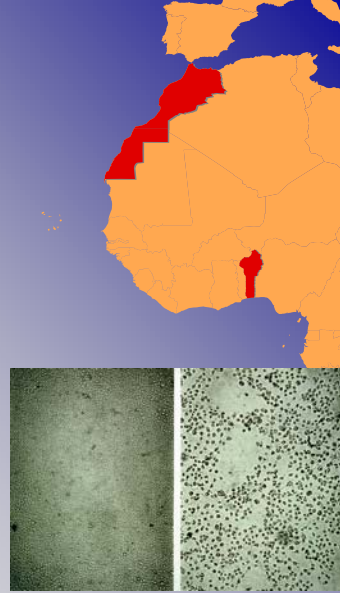
Unlike bacteria and parasites, they cannot replicate outside a host so their number decreases by time.

Viruses could be detected in sewage, mussels and water (river, pools, sea, wells).

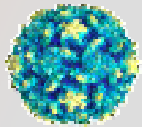




# Detectable viruses in environmental samples I



Enterovirus: Poliovirus, Coxsackievirus, Echovirus, Genome: RNA, 27nm

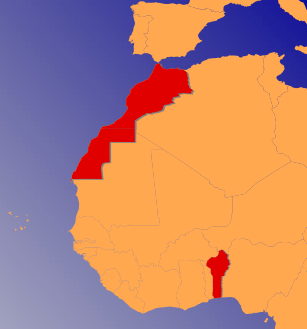


Incubation: 2-30 days

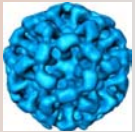
Symptoms: 99% asymptomatic

flu-like, malaise, meningitis, encephalitis

# Detectable Viruses in Environmental Samples II



## Norovirus:



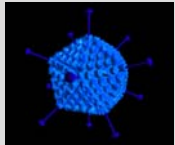
Genome: RNA, 27nm, 2 Genotypes (I-II)

Incubation: 1-12 h

Symptoms: Diarrhoea, vomit; (epidemical)



## Adenovirus:



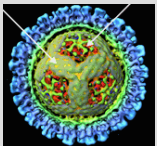
Genome: DNA, 70-100nm, Subtypes 49 (1-49)

Incubation: 2-7 days

Symptoms: Subtypes 40/41: Diarrhoea

Subtypes 8/3/7: Conjunctivitis

## Rotavirus:

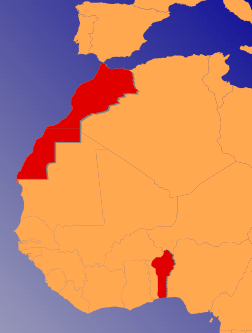


Genome: RNA, 76nm, 3 Genotypes (A-C)

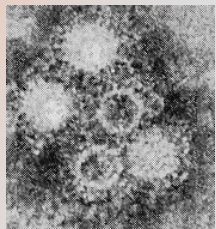
Incubation: 1-3 days

Symptoms: Diarrhoea (children)

# Detectable Viruses in Environmental Samples III



## Hepatitis-A-Virus:

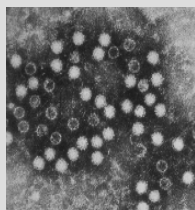


Genome: RNA, 28nm

Incubation: 14-50 days

Symptoms: abdominal pain, acute hepatitis,  
jaundice

## Hepatitis-E-Virus:

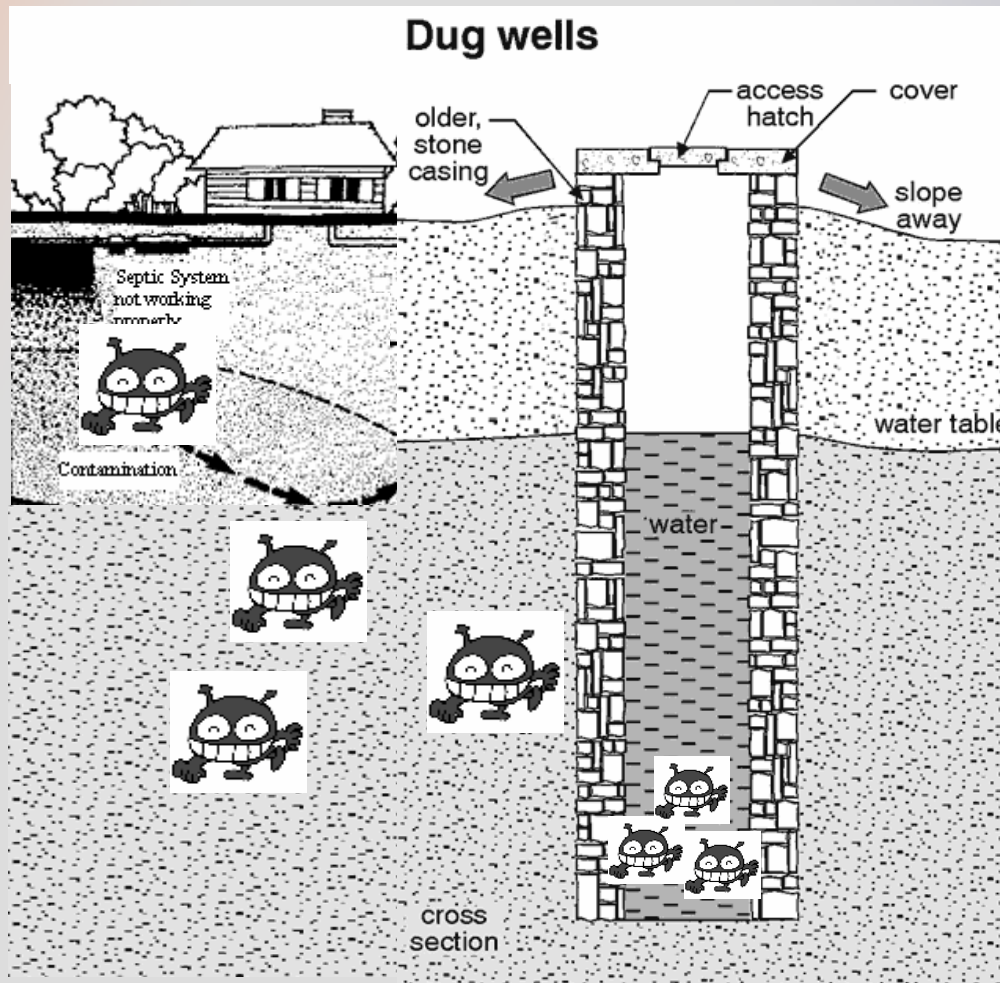
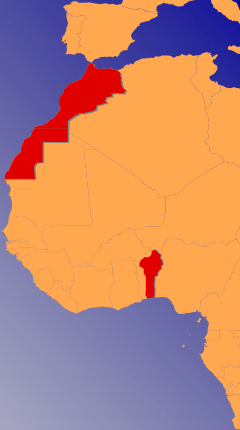


Genome: RNA, 32nm

Incubation: 28-50 days

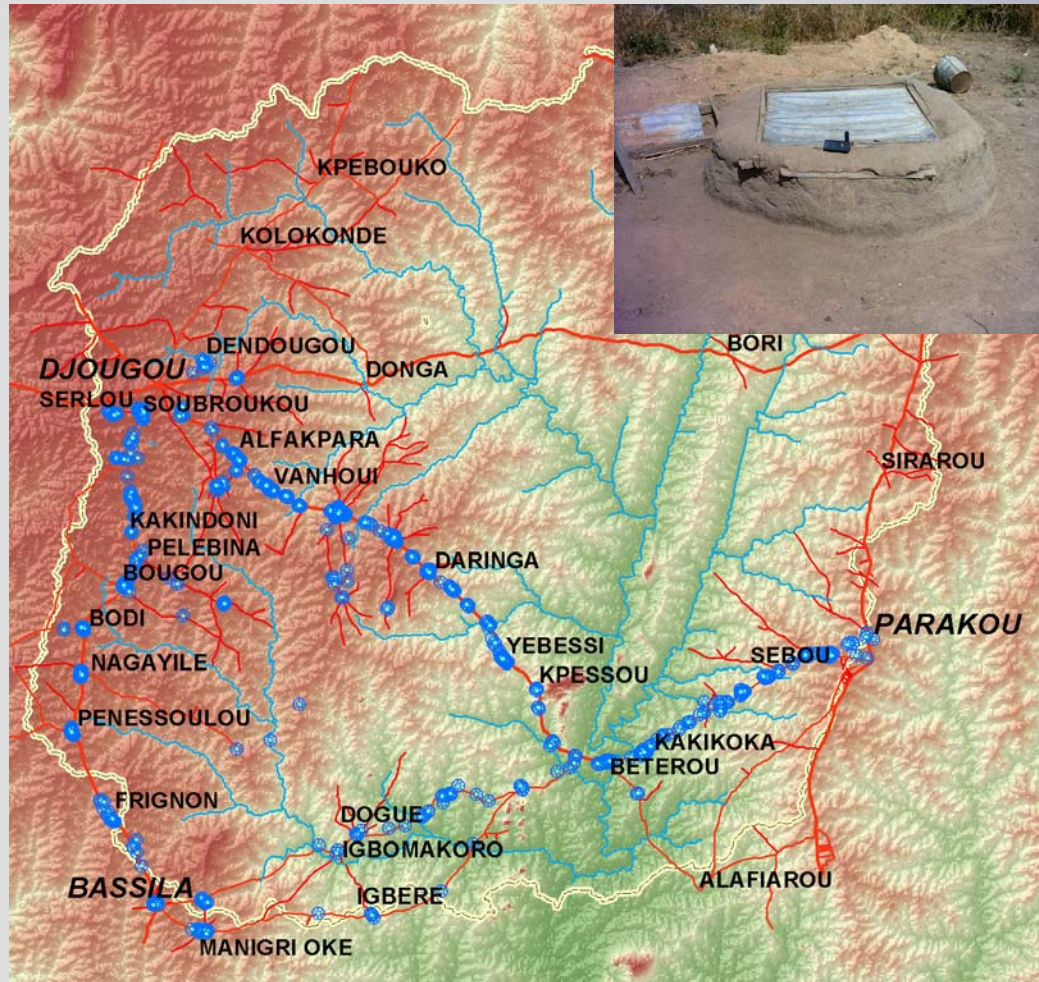
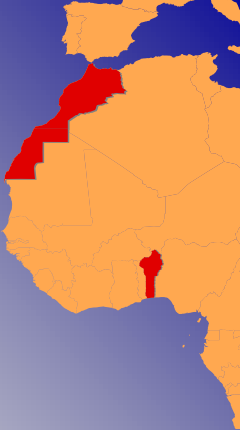
Symptoms: abdominal pain, acute hepatitis,  
jaundice,  
20% mortality in pregnant women

# Water wells



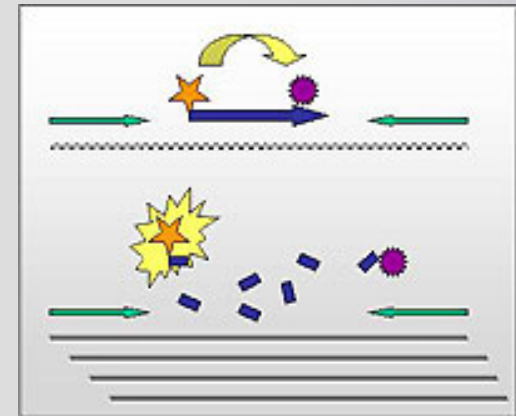
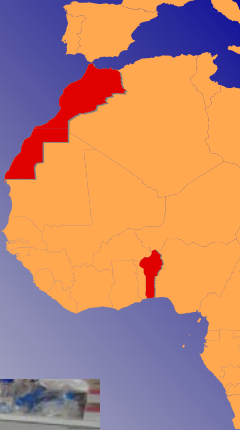


# *Wells - Database*



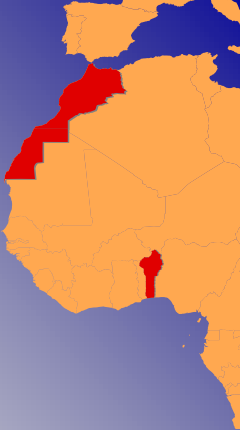


# Sample processing Benin - Cologne





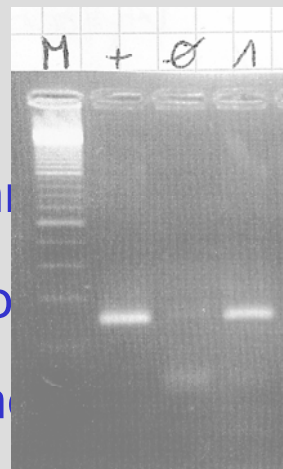
# Results I - water samples Benin I



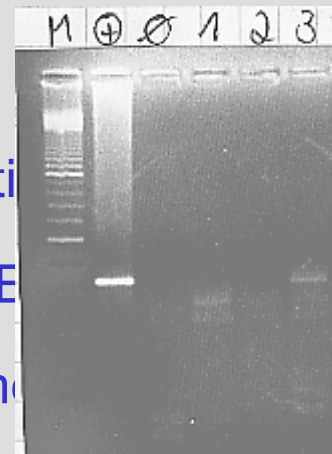
Enterovirus: 1/121 positive

Norovirus: 1/121 positive

Enterovirus



Norovirus

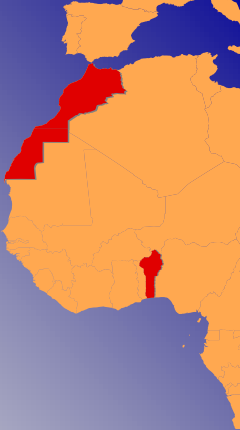


=> New PCR technique for qualitative detection of viruses in water by Real-time-PCR-technique

=> Routinely performed for water samples (E. coli, Enterococcus, etc.)

=> Extension of the detection spectrum by Real-time PCR-technique:

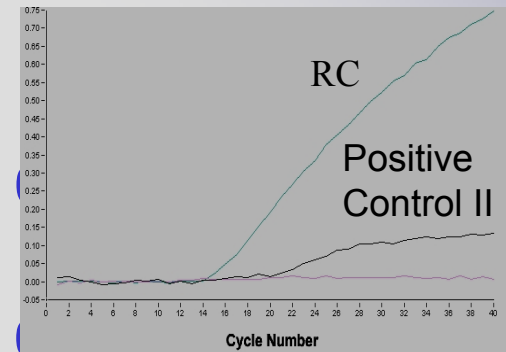
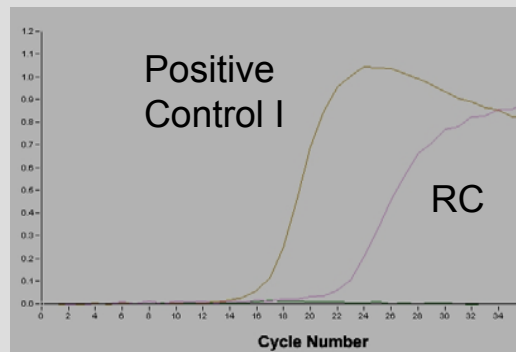
Hepatitis-A-Virus, Adenovirus, Rotavirus



# Results II - water samples / controls from Benin

Positive Controls:

Positive



HAV:

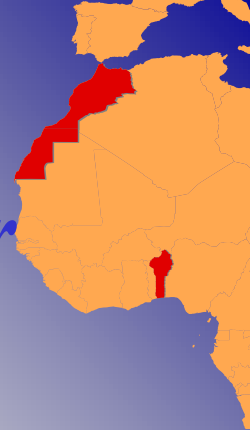
0/67 positive

Rotavirus:

in progress

## Results III –

# Validation of virus isolation from water



Adenoviruses were added to 10l water and analysed like water samples afterwards

adenovirus  
reference  
1ml with  
cop

added to  
10l H<sub>2</sub>O  
cop/ml

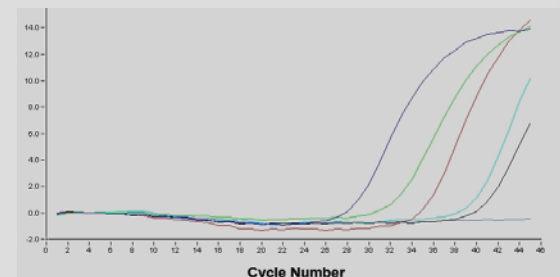
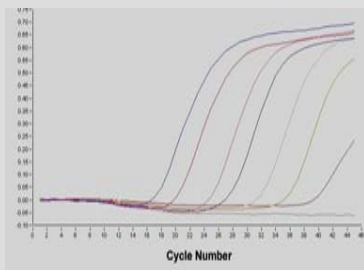
Concentration of  
detected Adenovirus  
cop/ml\*

\*at least 3 independent experiments

$3,3 \times 10^8$   
 $2,7 \times 10^7$   
 $3,3 \times 10^6$   
 $1,8 \times 10^5$

$3,3 \times 10^4$   
 $2,7 \times 10^3$   
 $3,3 \times 10^2$   
 $1,8 \times 10^1$

$2,6 \times 10^6$   
 $1,7 \times 10^5$   
 $1,9 \times 10^4$   
 $2,1 \times 10^3$



# Conclusions

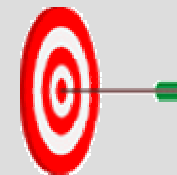
We established a system for virus detection in water samples from Benin approved by controls and validation (Benin/Cologne).



The low amount of positive samples could be explained with the temporary nature of virological contamination.



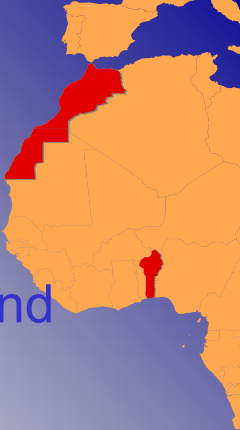
Identification and preferable investigation of high risk drinking water sources.







# *Perspectives*



## **A.**

Screening of stool samples from patients with diarrhoea for rota- and adenoviruses in local laboratories.

Investigation of drinking water sources used by positive patients and further symptomatic inhabitants

Analysis of water samples and genome sequencing of positive stool and water samples to investigate the infection chain.

In cooperation with  
R-Biopharm which provide 250 stool analysis kits for manufacturing costs  
Stratec NewGen, testing a new substance for transport and preservation

## **B.**

Sample collection in chosen villages with approved fecal contamination of wells in different seasonal settings





# *Thank you!!!!!!*

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Impetus Laboratory in Benin

Farouk Mazou  
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Martial Dossou

Et tous les collegues d'Impetus



University of Cologne



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